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Potential difference responses to nutrient K^+ , Na^+ and Cl^- changes with varying nutrient HCO_3^- in resting frog stomach

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The effects of changes in nutrient concentrations of K^+ , Na^+ and Cl^- on the transmucosal potential difference (PD) and resistance were compared for 25 and 5 mM nutrient HCO_3^- in resting fundus. With 25 mM HCO_3^- , increase of K^+ from 4 to 40 mM, decrease of Na^+ from 100 to 10 mM and decrease of Cl^- from 81 to 8.1 mM gave, 10 min after the change, ΔPD values of -23.2 , -15.1 and -21.3 mV, respectively. With 5 mM HCO_3^- , the same changes in nutrient ion concentration gave ΔPD values of -11.9 , -9.4 and -10.0 mV, respectively. From these results, in going from 25 to 5 mM HCO_3^- , it follows that the resistances of the ionic pathways for K^+ , Na^+ and Cl^- increased. The anomalous PD response following the increase in nutrient K^+ from 4 to 40 mM with 5 mM nutrient HCO_3^- gave further evidence that the resistance of the simple K^+ conductance pathway increased prior to the increase to 40 mM K^+ . The fact that 2 mM Ba^{2+} in the 25 mM HCO_3^- nutrient gave a smaller increase in resistance, compared to the decrease in nutrient HCO_3^- from 25 to 5 mM, supported the inference that resistances of ion pathways other than that of the K^+ pathway increased.

Introduction

Using ion substitution, in which an ion is replaced by a relatively impermeant ion, we found that there were conductance pathways for HCO_3^- in the nutrient membrane of the bullfrog antrum [1] and of the resting (or inhibited) fundus of *Rana pipiens* [2]. In contrast to the resting fundus, Sanders et al. [3] found in the secreting fundus no significant change in potential difference (PD) with a change in nutrient HCO_3^- concentration and hence had no evidence of HCO_3^- conductance pathways in the nutrient membrane. We note further that, previously, Flemstrom and Sachs [4] reported a change in PD resulting from a change in HCO_3^- concentration both in the nutrient and secretory sides of the antrum. They did not determine, however, whether a linear relationship existed between $|\Delta PD|$ and the log of the HCO_3^- concentration. We reported such a relationship for both the antrum [1] and the resting fundus [2].

Moreover, with a decrease in HCO_3^- concentration in the nutrient solution, there was an increase in transmucosal resistance. It is difficult to explain the linear

relationship when the resistance increases with a decrease in HCO_3^- concentration. An explanation which we gave assumed that not only did the resistance of the HCO_3^- pathway vary, but also the resistances of other ionic pathways varied inversely with the HCO_3^- concentration (see Refs. 1,2 and Discussion).

The purpose of the present paper was to determine which of the known conductance pathways (e.g., K^+ , Na^+ , Cl^-) other than the HCO_3^- pathway were involved in the increase in resistance.

Methods

Experiments were performed on fundi of stomachs of *R. pipiens* by an in vitro method in which the stomachs were mounted between a pair of cylindrical chambers [5]. All experiments began with standard Cl^- solutions on both sides of the mucosa. The Cl^- nutrient (serosal) solution contained (in mM): Na^+ , 102; K^+ , 4; Ca^{2+} , 1; Mg^{2+} , 0.8; Cl^- , 81; HCO_3^- , 0.8; $H_2PO_4^-$, 25; phosphate, 1; and glucose, 10; and the Cl^- secretory (mucosal) solution, which is hypertonic [6], contained Na^+ , 156; K^+ , 4; and Cl^- , 160. In the decrease in nutrient HCO_3^- from 25 to 5 mM, standard nutrient solutions which were phosphate-free were used for both HCO_3^- concentrations. In the decrease from 25 to 5 mM HCO_3^- , SO_4^{2-} replaced HCO_3^- and sucrose was added to main-

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tain osmolality. Various phosphate-free modifications were also used in which the HCO_3^- concentrations were reduced from 25 mM HCO_3^- . In all these solutions, the Ca^{2+} , Mg^{2+} , K^+ and Cl^- concentrations were unaltered unless a specific experiment demanded a change in K^+ or Cl^- concentration. For increases in K^+ concentration on the nutrient side, K^+ replaced Na^+ and, for decreases in Na^+ concentration on the nutrient side, choline replaced Na^+ or Mg^{2+} replaced Na^+ . For a decrease in Cl^- concentration, SO_4^{2-} replaced Cl^- and sucrose was added to make up any osmotic deficit.

In these studies, the transmucosal resistance, the transmucosal PD and the H^+ secretory rate were measured. Two pairs of electrodes were used, one for sending current across the mucosa and the other for measuring the PD. The PD is considered positive when the nutrient side is positive relative to the secretory side of the stomach. The resistance was determined as the change in PD per unit of applied current. Current (20 μA per 1.3 cm^2 of tissue area) was applied for 1 or 2 s, first in one direction and 2 or 3 s later, in other direction. The H^+ secretory rate was determined by the pH-stat method of Durbin and Heinz [7]. The pH of the secretory solution was generally maintained between 4.7 and 5.0. Moreover, both sides of the gastric mucosa were gassed with 95% O_2 /5% CO_2 throughout these experiments and 0.1 mM histamine in the nutrient solution was used to stimulate secretion. For inhibition, 20 mM SCN^- in the secretory solution decreased the H^+ secretory rate to zero.

In previous studies on the nutrient side, due to the existence of a diffusion barrier between the nutrient solution and the nutrient membrane, it took about 10 min (approx. 5 time constants) for the concentration of the ion at the cell membrane to attain the new concentration in the nutrient solution [8]. Hence the PD

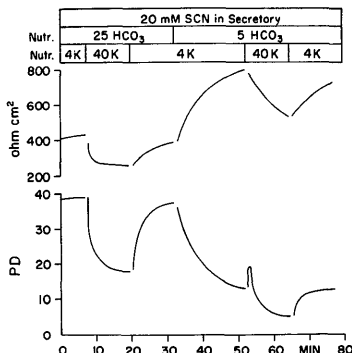


Fig. 1. Effect of changes in K^+ concentration on the nutrient side from 4 to 40 mM and back to 4 mM with 25 and 5 mM nutrient HCO_3^- and with 20 mM SCN^- in the secretory side. Resistance and PD are plotted vs. time. Concentrations are in mM.

and resistance were generally read at the 10 min mark following the change to the new solution.

Results

PD responses and resistance changes of the resting fundus due to changes in K^+ concentration in nutrient solutions with 25 and with 5 mM HCO_3^-

Fig. 1 is a plot of transmucosal resistance and PD versus time for the change from 4 to 40 mM K^+ and back to 4 mM K^+ for resting fundus in the presence of

TABLE I

Effect on PD and resistance of changes in K^+ concentrations in the nutrient solution containing varying concentrations of HCO_3^-

Values are means \pm S.D. Student's *t*-test using paired observations was used to determine the level of significance. Columns labelled PD and *R* refer to the control values of transmucosal potential difference and corresponding resistance and columns labelled ΔPD_M , ΔPD and ΔR refer respectively to changes of the PD at the maximum and at 10 min and the change in *R* at 10 min following the change to the final concentration of the ion. Number of experiments, 6.

[K ⁺] (mM)		PD (mV)	ΔPD_M (mV)	ΔPD (mV)	<i>R</i> ($\Omega \cdot \text{cm}^2$)	ΔR ($\Omega \cdot \text{cm}^2$)
orig. soln.	final soln.					
Resting state: 25 mM nutrient HCO ₃ ⁻						
4	40	42.9 ± 9.2	–	–23.2 ± 5.9 *	335 ± 63	–119 ± 35 *
40	4	18.1 ± 4.3	–	20.6 ± 3.6 *	205 ± 30	53 ± 39 ^b
Resting state: 5 mM nutrient HCO ₃ ⁻						
4	40	20.6 ± 8.2	4.9 ± 2.9 **	–11.9 ± 4.3 *	680 ± 200	–192 ± 29 *
40	4	7.0 ± 3.5	–	9.4 ± 4.5 *	478 ± 123	188 ± 110 *

* $P < 0.01$;

b $P < 0.02$;

* ΔPD_M is the average of five experiments, since no maximum was obtained in one experiment.

TABLE II

Effect on PD and resistance of changes in Na^+ concentrations in the nutrient solution containing varying concentrations of HCO_3^-

See Table I for details. Number of experiments, 6.

[Na ⁺] (mM)		PD (mV)	Δ PD (mV)	R ($\Omega \cdot \text{cm}^2$)	ΔR ($\Omega \cdot \text{cm}^2$)
orig. soln.	final soln.				
Resting state: 25 mM HCO ₃ ⁻					
100	10	41.7 ± 7.9	-15.1 ± 3.1 ^a	381 ± 70	180 ± 57 ^a
10	100	25.2 ± 6.5	15.5 ± 4.4 ^a	590 ± 132	-156 ± 61 ^a
Resting state: 5 mM HCO ₃ ⁻					
100	10	21.4 ± 5.7	-9.4 ± 2.0 ^a	755 ± 93	132 ± 199
10	100	8.0 ± 2.9	5.9 ± 2.3 ^a	839 ± 283	-51 ± 91

^a $P < 0.01$.

25 mM HCO_3^- and later in the presence of 5 mM HCO_3^- . With 25 mM HCO_3^- , the increase in K^+ from 4 to 40 mM in the nutrient solution decreased the resistance and decreased the PD. With the change to 5 mM HCO_3^- , the resistance increased substantially. Then, in the change from 4 to 40 mM K^+ , there was a biphasic response: an initial increase in PD, an anomalous response, previously attributed to the existence of a Na^+/K^+ -ATPase pump [9-11], followed by a decrease in PD below the control level, a normal response but of lesser magnitude than with 25 mM HCO_3^- . Here also the resistance decreased with the change from 4 to 40 mM K^+ . These results may be explained by an increase in resistance of the simple K^+ resistance pathway with 4 mM K^+ in the nutrient solution due to the decrease in the concentration of nutrient HCO_3^- (see Discussion). The return to 4 mM K^+ brought the PD and resistance back toward control levels at 25 and 5 mM nutrient HCO_3^- .

In Table I, in going from 4 to 40 mM K^+ , Δ PD at 10 min was -23.2 mV with 25 mM nutrient HCO_3^- and -11.9 mV with 5 mM nutrient HCO_3^- . In five of the six experiments, the PD response was biphasic in that a maximum PD occurred generally within 1 min after the K^+ increase from 4 to 40 mM was made and the maximum was followed by a decrease in PD below the

control level. The maximum Δ PD did not occur in one of the six experiments and the average value of the maximum Δ PD for the five experiments was 4.9 mV. In the case of the antrum, the maximum Δ PD was about 8 mV [11]. The respective decreases in resistance for 25 and 5 mM nutrient HCO_3^- were 119 and 192 $\Omega \cdot \text{cm}^2$. In percentages, the decreases were respectively 36 and 28%. Upon return to 4 mM K^+ , only a normal response was observed, that is, the PD and resistance both increased towards control values.

PD responses and resistance changes of the resting fundus due to changes in the Na^+ concentration in nutrient solutions with 25 and with 5 mM HCO_3^-

Table II shows results for changes in Na^+ concentration in resting fundus similar to that obtained above for changes in K^+ concentration. For decreases in nutrient Na^+ concentration from 100 to 10 mM for 25 and 5 mM HCO_3^- , the respective decreases in PD were 15.1 and 9.4 mV. Since the PD decreases with a decrease in nutrient Na^+ concentration, this result was previously considered as an anomalous response and was attributed to the existence of an electrogenic NaCl symport in the nutrient membrane [12,13]. The increase in resistance was significant only with 25 mM HCO_3^- and was in this case 180 $\Omega \cdot \text{cm}^2$. Upon the return to 100 mM

TABLE III

Effect on PD and resistance of changes in Cl^- concentrations in the nutrient solution containing varying concentrations of HCO_3^-

See Table I for details. Number of experiments, 6.

[Cl ⁻] (mM)		PD (mV)	ΔPD (mV)	R (Ω·cm ²)	ΔR (Ω·cm ²)
orig. soln.	final soln.				
Resting state: 25 mM HCO ₃ ⁻					
81	8.1	45.2 ± 3.0	-21.3 ± 2.6 ^a	309 ± 90	71 ± 97
8.1	81	23.1 ± 2.7	21.4 ± 2.5 ^a	381 ± 154	-18 ± 57
Resting state: 5 mM HCO ₃ ⁻					
81	8.1	25.2 ± 7.8	-10.0 ± 2.7 ^a	688 ± 196	-4 ± 62
8.1	81	9.8 ± 8.8	8.5 ± 3.7 ^a	713 ± 199	69 ± 67

^a $P < 0.01$.

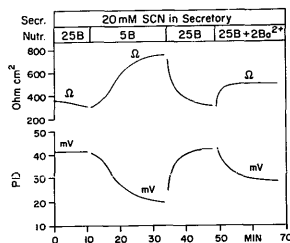


Fig. 2. Effect of change in nutrient HCO_3^- from 25 to 5 mM and back to 25 mM followed by effect of addition of 2 mM Ba^{2+} to the 25 mM nutrient solution. Throughout 20 mM SCN^- was present in the secretory side. Resistance and PD are plotted vs. time. Concentrations are in mM and B refers to HCO_3^- .

Na^+ , the results returned to control values for 25 mM HCO_3^- but only partly for 5 mM HCO_3^- .

PD responses and resistance changes of the resting fundus due to changes in the Cl^- concentration in nutrient solutions with 25 and with 5 mM HCO_3^-

Table III shows the results for decreases in Cl^- concentration in the nutrient solution from 81 to 8.1 mM for 25 and for 5 mM nutrient HCO_3^- . The respective decreases in PD were 21.3 and 10.0 mV. The changes in resistance were, however, not significant. Upon return to 8.1 mM Cl^- , the PD returned to control or near control values.

Effect on PD and resistance due to a decrease in nutrient HCO_3^- from 25 to 5 mM HCO_3^- compared to the addition of 2 mM Ba^{2+} to the 25 mM nutrient HCO_3^- solution

It has long been established that Ba^{2+} in the nutrient solution of frog gastric mucosa blocks the K^+ conductance pathway [14,15] (see Discussion). It was therefore thought that, if the resistance increase due to Ba^{2+} was about the same as the resistance increase due to the

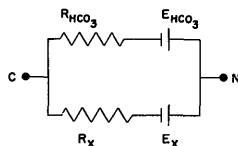


Fig. 3. Equivalent circuit for nutrient membrane consisting of a HCO_3^- limb and an X limb representing all other ionic pathways. C and N refer respectively to cell and nutrient.

decrease in HCO_3^- , then only the resistance of the K^+ pathway increased due to a decrease in HCO_3^- . In reality, as described below, the decrease in HCO_3^- gave a greater increase in resistance than the addition of Ba^{2+} to a 25 mM HCO_3^- nutrient solution. Hence this result gave further evidence that other pathways like Na^+ and Cl^- also increased in resistance with a decrease in HCO_3^- .

Fig. 2 shows the results under the two circumstances, a decrease in nutrient HCO_3^- and the addition of nutrient Ba^{2+} . In the first case, the resistance increased markedly and the PD decreased markedly. In the second case, similar changes occurred in resistance and PD but to a lesser extent.

In Table IV, the decrease in nutrient HCO_3^- from 25 to 5 mM decreased the PD by 17.7 mV and increased the resistance by $284 \Omega \cdot \text{cm}^2$, while the addition of Ba^{2+} to 2 mM in the nutrient solution decreased the PD by 10.0 mV and increased the resistance by $112 \Omega \cdot \text{cm}^2$. The respective increases in resistance were, in percentages, 72 and 28%. It is to be noted that the relatively low increase in resistance ($112 \Omega \cdot \text{cm}^2$) due to the addition of Ba^{2+} is to be expected in the resting fundus, whereas much larger increases in resistance (about $700 \Omega \cdot \text{cm}^2$ [16]) occur for stomachs with high acid-secreting rates. As expected [14], the addition of Ba^{2+} to 2 mM in the 25 mM HCO_3^- nutrient solution gave essentially maximal increase in resistance. In the present experiments, with 5 mM HCO_3^- in the nutrient solution, the addition of Ba^{2+} in steps up to 6 or 8 mM

TABLE IV

Effect on PD and resistance for a decrease in nutrient HCO_3^- and for the addition of nutrient Ba^{2+}

See Table I for details. Number of experiments, 6.

[HCO ₃ ⁻] (mM)		PD (mV)	ΔPD (mV)	R (Ω·cm ²)	ΔR (Ω·cm ²)
orig. soln.	final soln.				
Case I: No Ba ²⁺ in original or final nutrient solution					
25	5	36.5 ± 7.4	-17.7 ± 2.8 ^a	393 ± 145	284 ± 94 ^a
Case II: 2 mM Ba ²⁺ in final nutrient solution					
25	25	36.8 ± 5.8	-10.0 ± 1.5 ^a	403 ± 161	112 ± 28 ^a

^a $P < 0.01$.

gave no further increase in resistance, and at higher concentrations a small decrease on occasion.

Discussion

As mentioned in the Introduction, it is difficult to explain the linear relationship between ΔPD and the log of the HCO_3^- concentration with an increase in transmucosal resistance. To see how this difficulty arises, let us refer to Fig. 3 in which is shown an electrical circuit consisting of a HCO_3^- limb and an X limb for all other ion pathways. Each limb consists of a resistance, R , and an emf, E . Even though the transport system is complex in the gastric mucosa, one can use a simplified circuit for the purpose under discussion. For this circuit, it follows that the PD is given by

$$PD = (R_X E_{HCO_3^-} + R_{HCO_3^-} E_X) (R_{HCO_3^-} + R_X)^{-1}$$

If R_X and $R_{HCO_3^-}$ were constant, then for a change in HCO_3^- concentration, it would follow (since $\Delta E_X = 0$) that

$$\Delta PD = R_X (R_{HCO_3^-} + R_X)^{-1} \Delta E_{HCO_3^-}$$

and a linear relationship would exist between ΔPD and $\Delta E_{HCO_3^-}$, and hence between ΔPD and the log of the HCO_3^- concentration. However, since changes in HCO_3^- concentration gave rise to changes in transmucosal resistance, some hypothesis other than constancy of R_X and $R_{HCO_3^-}$ was needed. As a possibility, it was assumed that, if R_X and $R_{HCO_3^-}$ both varied inversely with the HCO_3^- concentration, then $R_X (R_{HCO_3^-} + R_X)^{-1}$ would remain constant. In general, it would be very difficult to explain the linear relationship based on a model in which either resistance alone of the parallel pathways changed with variation in the HCO_3^- concentration.

The present study provided a means of determining whether ionic pathways besides the HCO_3^- pathway underwent a change in resistance with a change in HCO_3^- concentration. Such information is associated with the result that ΔPD due to a change in concentration of an ion (e.g., K^+ , Na^+ , Cl^-) in the nutrient is less with a low nutrient concentration of HCO_3^- than with a high concentration of HCO_3^- . The decrease in ΔPD results generally from an increase in resistance of the ionic pathway under consideration. In addition, the 10-fold increase in K^+ in the nutrient solution containing 5 mM HCO_3^- gave an initial transient increase in ΔPD with a maximum of 4.9 mV. Such transient increases in PD were not observed with 25 mM HCO_3^- .

As stated above, the PD responses provide some idea of the changes in conductance in going from 25 to 5 mM HCO_3^- in the nutrient solution. The K^+ transport is associated with both a simple conductance pathway and a Na^+/K^+ -ATPase pump pathway [8–11]. Since

the resistance of the latter pathway is normally high, changes in ΔPD often reflect changes in the resistance of the simple K^+ conductance pathway. However, there are exceptions [9–11], as occurred with 5 mM HCO_3^- in the 4 mM K^+ nutrient solution. In this case, the resistance of the simple K^+ conductance pathway increased so that the PD response from an increase in K^+ was due to the Na^+/K^+ -ATPase pump, that is, the increase from 4 to 40 mM K^+ gave an anomalous PD response. The presence of 40 mM K^+ in the nutrient solution decreases the resistance of the simple K^+ conductance pathway. For this reason, the anomalous response was quickly followed by a normal response (for details, see Ref. 10).

As for other pathways, the Na^+ transport is associated mainly with the NaCl symport pathway [12,13]. Then for Na^+ , a decrease in magnitude in ΔPD due to a decrease in HCO_3^- concentration reflects essentially an increase in the resistance of the symport pathway. For Cl^- , there are two pathways: a simple conductance pathway and the NaCl symport pathway [8,12,13]. It would be difficult to determine which pathway or whether both pathways are affected by a decrease in nutrient HCO_3^- . The results with Ba^{2+} support the increase in resistance in pathways besides the K^+ pathway.

Evidence that Ba^{2+} in the nutrient solution blocks the nutrient K^+ pathway and increases markedly the resistance of this pathway is provided by the following information and experiment. In the absence of Ba^{2+} , we note that, in the secretory membrane, the resistance of the K^+ pathway is high and that of the Cl^- pathway is low [5,17] while, in the nutrient membrane, the resistance of the K^+ pathway is low and that of the Cl^- pathway(s) about twice that of the K^+ pathway [8]. As a consequence, the ratio of the K^+ resistance to the Cl^- resistance in the secretory membrane is considerably greater than the ratio of the K^+ resistance to the Cl^- resistance in the nutrient membrane. Then, if an external step current is sent through the frog gastric mucosa, the transient voltage response versus time has as one of its components an exponential curve with a very long time constant [17]. If, however, Ba^{2+} is added to the nutrient solution, the above-mentioned ratios approach equality and the transient voltage-response versus time is immediate [17]. Thus this experiment, in addition to the one previously mentioned [15], proves that Ba^{2+} in the nutrient solution increases the resistance of the K^+ pathway markedly. In the present study, it becomes evident from these considerations that any further increase in transmucosal resistance, such as that resulting from the decrease in nutrient HCO_3^- , must arise from the increase in resistance in pathways other than the K^+ pathway.

We wish now to comment briefly on intracellular pH regulation. Paradiso et al. [18] found evidence for neu-

tral Na^+-H^+ and Cl^--OH^- (HCO_3^-) exchange in gastric glands of rabbit stomach. These mechanisms were associated with internal pH regulation. In the intact frog gastric mucosa there is considerable evidence for conductance pathways. From the present study in the resting fundus we might infer that the decrease in nutrient HCO_3^- decreases internal pH. It would appear then that, while ions like Na^+ and Cl^- play a role in pH regulation, the pH in turn plays a role in controlling the resistance of ion pathways such as those for K^+ , Na^+ and Cl^- transport.

Acknowledgments

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References

- Schwartz, M., Carrasquer, G. and Rehm, W.S. (1985) *Biochim. Biophys. Acta* 816, 251–257.
- Schwartz, M., Carrasquer, G. and Rehm, W.S. (1985) *Biochim. Biophys. Acta* 819, 187–194.
- Sanders, S.S., O'Callaghan, J., Butler, C.F. and Rehm, W.S. (1972) *Am. J. Physiol.* 222, 1348–1354.
- Flemstrom, G. and Sachs, G. (1975) *Am. J. Physiol.* 228, 1188–1198.
- Rehm, W.S. (1962) *Am. J. Physiol.* 203, 63–72.
- Rehm, W.S., Chu, T.C., Schwartz, M. and Carrasquer, G. (1983) *Am. J. Physiol.* 245 (Gastrointest. Liver Physiol. 8), G143–G156.
- Durbin, R.P. and Heinz, E. (1959) *J. Gen. Physiol.* 41, 1035–1047.
- Spangler, S.G. and Rehm, W.S. (1968) *Biophys. J.* 8, 1211–1227.
- Schwartz, M., Chu, T.C., Carrasquer, G. and Rehm, W.S. (1981) *Biochim. Biophys. Acta* 649, 253–261.
- Schwartz, M., Kissel, D.E., Carrasquer, G. and Rehm, W.S. (1983) *Biochim. Biophys. Acta* 727, 45–55.
- Schwartz, M., Carrasquer, G. and Rehm, W.S. (1984) *Biochim. Biophys. Acta* 769, 105–116.
- Carrasquer, G., Chu, T.C., Rehm, W.S. and Schwartz, M. (1982) *Am. J. Physiol.* 242 (Gastrointest. Liver Physiol. 5), G620–G627.
- Carrasquer, G., Kissel, D.E., Rehm, W.S. and Schwartz, M. (1983) *Am. J. Physiol.* 245 (Gastrointest. Liver Physiol. 8), G559–G561.
- Schwartz, M., Pacifico, A.D., MacKrell, T.N., Jacobson, A. and Rehm, W.S. (1968) *Proc. Soc. Exp. Biol. Med.* 127, 223–225.
- Pacifico, A.D., Schwartz, M., MacKrell, T.N., Spangler, S.G., Sanders, S.S. and Rehm, W.S. (1969) *Am. J. Physiol.* 215, 536–541.
- Sanders, S.S., Shoemaker, R.L. and Rehm, W.S. (1977) *Am. J. Physiol.* 233 (Endocrinol. Metab. Gastrointest. Physiol. 2), E298–E307.
- Kidder, G.W., III and Rehm, W.S. (1970) *Biophys. J.* 10, 215–236.
- Paradiso, A.M., Negulescu, P.A. and Machen, T.E. (1986) *Am. J. Physiol.* 250 (Gastrointest. Liver Physiol. 13), G524–G534.